

# MONT

## Montana Nanotechnology Facility

*An NSF NNCI Node in the Northern Rocky Mountain Region*



### Year 1 Activities Snapshot



David Dickensheets

NNCI Annual Conference, January 17-18, 2017

# Our Team:



David  
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Recep Avci  
ICAL



Phil Stewart  
CBE



Mark Young  
CBIN / VPR



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Carleton College  
Science Education  
Resource Center



Dave Mogk  
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ICAL Manager



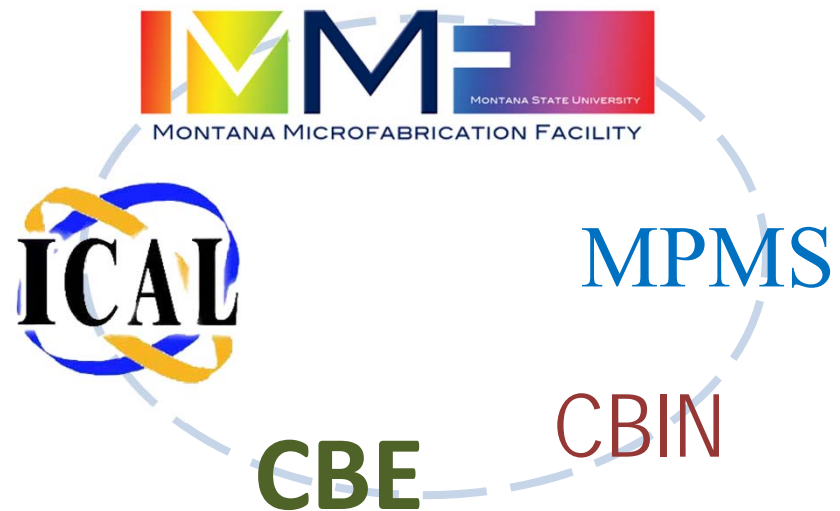
# Our Vision

The Montana Nanotechnology Facility (MONT) promotes nanotechnology discovery, education and outreach by providing access to shared-use instruments, expert training on their safe and effective use, and broad-based education about nanoscale science and technology for teachers and learners who come from diverse communities.

# Overview and Core Facilities

Coordinated access to and training on shared equipment housed in 5 campus facilities:

- Montana Microfabrication Facility
- Imaging and Chemical Analysis Lab
- Center for Biofilm Engineering
- Center for Bio-Inspired Nanomaterials
- Metabolomics, Proteomics and Mass Spectroscopy facility



# Core Fabrication



CBE

- Processes set up for 4 inch wafers and pieces
  - Wet benches
  - Spinners – pos, neg photoresists, ebeam resists, lots of SU-8 work
  - Patterning – contact litho; FE-SEM with NPGS, Dip-Pen nanowriting
  - Thermal processing – oxidation and anneal; solid source diffusion
  - PVD - Angstrom Engineering sputter, e-beam evap, thermal evap
  - Dry etching – cryo ICP; RIE; O2 ash;



# Core Characterization

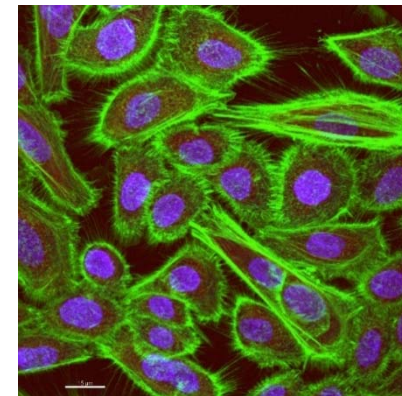
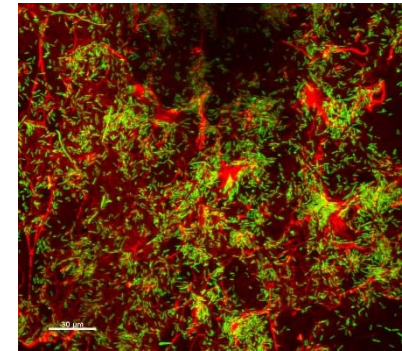
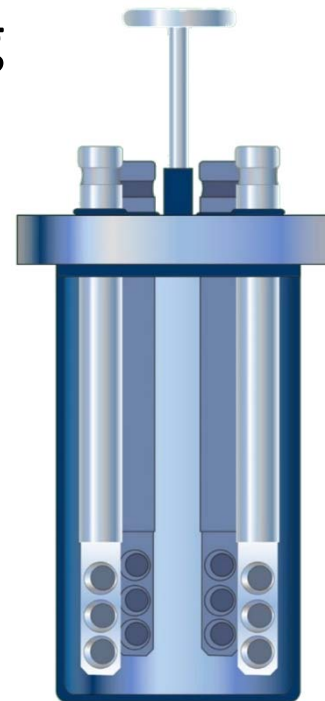


- **Thin films:** Ambios profilometer, Nanospec, resistivity probes
- **Device characterization:** probe stations, SPA, wire bonders
- **Imaging:** Metrology and stereo microscopes, fluorescence microscopes, SEMs, AFMs, TEM
- **Spectroscopy:** Auger, TOF-SIMS, XPS, EBSD, EDX, and Mass Spec capabilities spanning small to large molecules

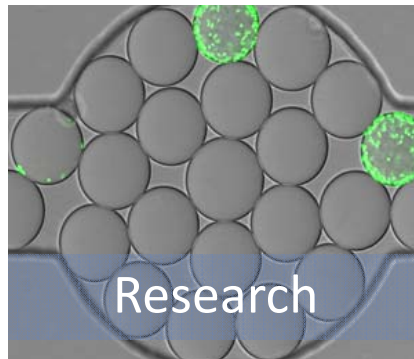


# CBE CBIN Unique Capabilities for Bio + Nano

- Center for Biofilm Engineering
  - Incubation capabilities for biofilms, mammalian cells and thermophiles
  - Chromatography and spectroscopy tools
  - Genetic sequencing
  - Advanced confocal and fluorescence imaging – live cell imaging



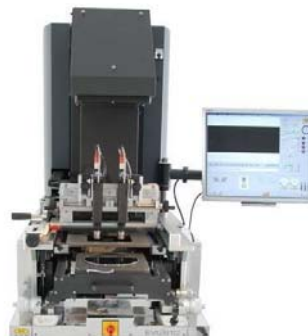
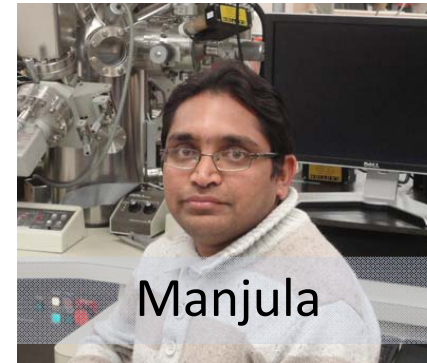
# Year 1 Highlights





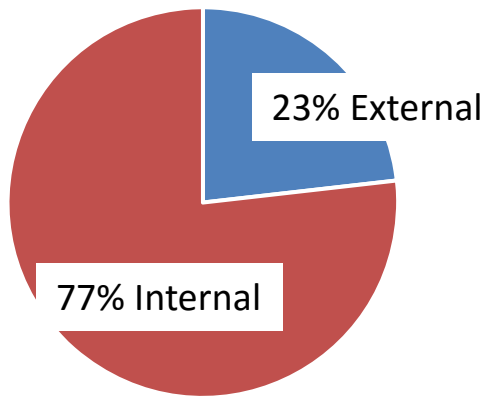
# Facility Enhancement

- Added Staff: litho/microfluidics, characterization, user liaison
- New Equipment: NanoAuger (PHI 710), EVG aligner, Filmetrics

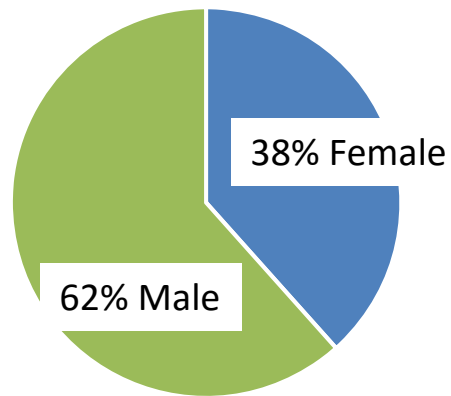


# Users

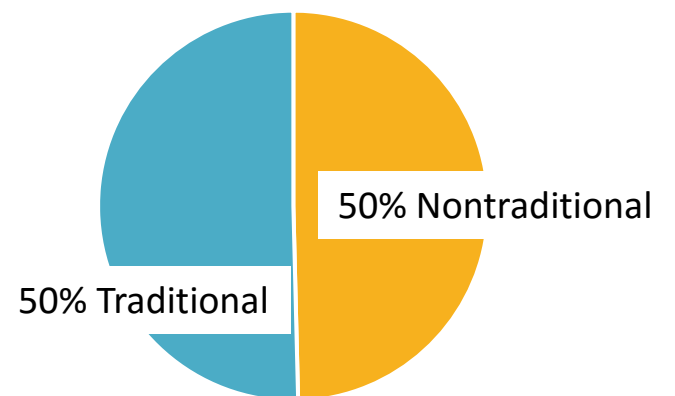
125 Total Users in Y1



External Users

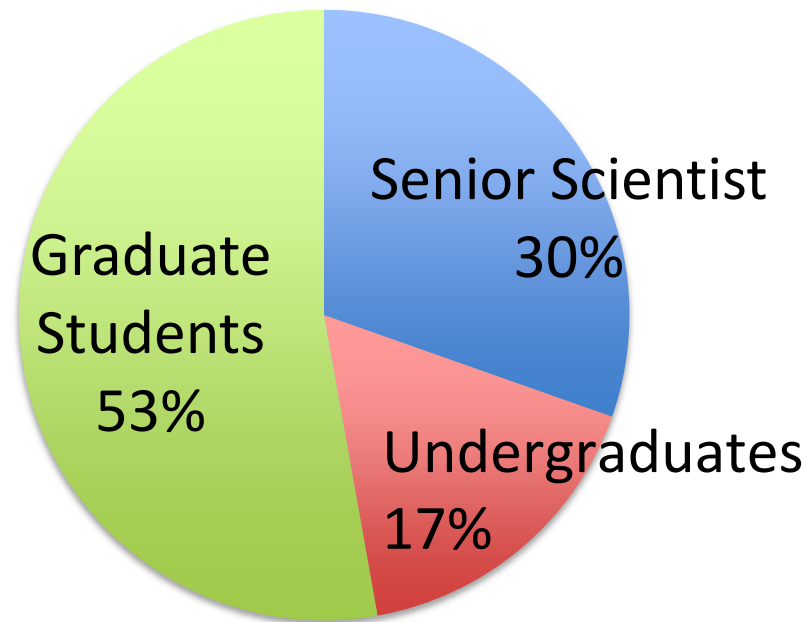


Gender



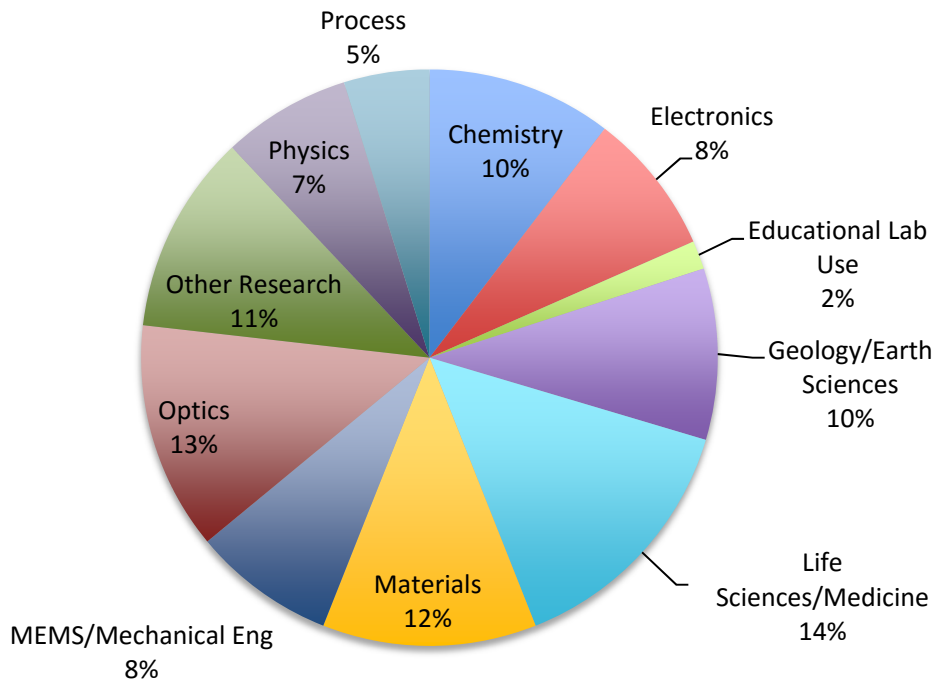
Non-Traditional

## Users – Academic Rank

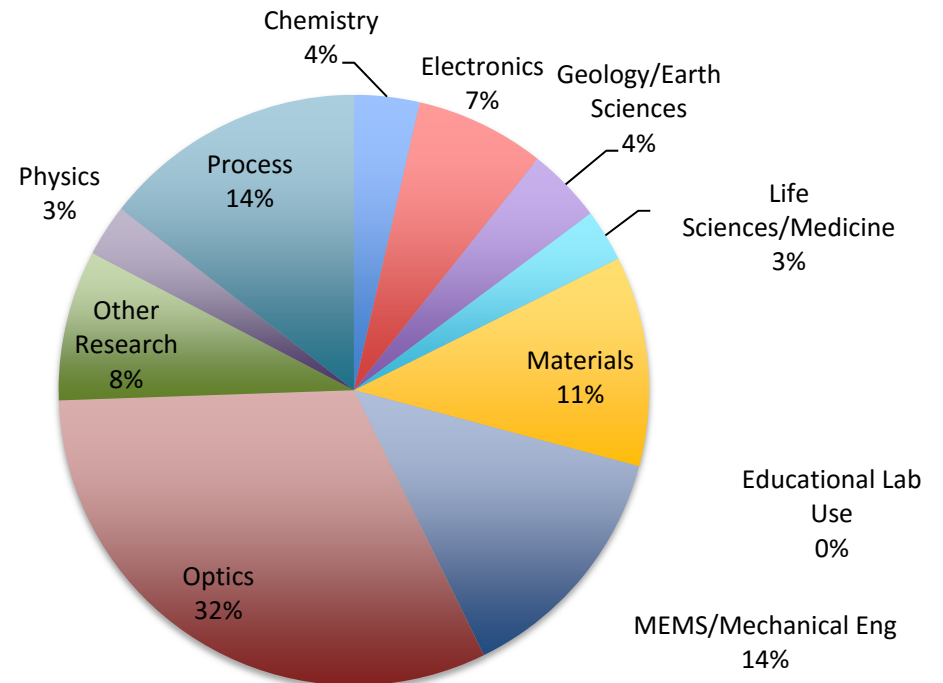


# Users – Discipline Breakdown

## Users by discipline



## Fees collected by discipline



# User-Driven New Directions

Proposals pending for new space and new equipment to support “Soft Lithography” and 3D printing for microfluidics research and training.

# Education and Outreach

- **70 undergraduates** used MONT for courses on Microfabrication and Photovoltaics
- Photovoltaics summer course for **secondary science teachers** (>60 so far)
- **Summer REU** in 2017, cooperatively with ECE department
- **Industry workshop**: Nanotech for Biofilms
- **Webinars**
- **K-12 Outreach**: Nanodays, Shadow an Engineer



**MONT: Montana Nanotechnology Facility Webinar Series**

### Imaging Microorganisms on Surfaces

**Presenters:** Philip S. Stewart and Betsy Pitts  
**Center for Biofilm Engineering  
Montana State University**

**December 7, 2016**

**Logistics**  
**Date and Time:** December 7, 2016, 1:30 to 2:30 pm (Mountain Standard Time)  
**Registration:** Registration is free, but we do need participants to pre-register using the registration form.

Please register by Friday, December 2, using this [Registration Form](#); seating is limited to 30 in Barnard Hall and 100 distance-videlink sites.

There are two options to participate:

- Attend in person: Burns Communication Center, Barnard Hall Room 127, Montana State Univ.
- Attend by videolink. An access code will be sent prior to the webinar to give you access to Adobe Connect.

**ABSTRACT:** Webinars on confocal microscopy techniques

This webinar will discuss the use of confocal microscopy to study microorganisms on engineered surfaces. At Montana State University, we have a wide range of confocal microscopes and imaging systems that are accessible to all users through the Montana Nanotechnology Facility on the same campus. A few examples of the types of systems that are accessible to users through the Montana Nanotechnology Facility on the same campus. A few examples of the types of systems that are accessible to users through the Montana Nanotechnology Facility on the same campus. A few examples of the types of systems that are accessible to users through the Montana Nanotechnology Facility on the same campus.



# Education and Outreach

- In cooperation with Carleton College Science Education Resource Center (SERC):

## Web-based educational and instructional resources

- Important nanotechnology processes (<http://serc.carleton.edu/18410>)
- Nanotechnology applications
- Ethics and societal impacts of nanotechnology
- Teaching resources – integrating discussion of ethics and society into all of our educational activities

<http://serc.carleton.edu/geoethics/index.html>

## – Web-based outreach

- <http://serc.carleton.edu/8640>

## Time-of-Flight Secondary Ion Mass Spectrometry (ToF-SIMS)

David W. Mogk, Montana State University

### Time-of-Flight Secondary Ion Mass Spectrometry (ToF-SIMS)

Time-of-Flight Secondary Ion Mass Spectrometry (ToF-SIMS) is a surface-sensitive analytical method that uses a pulsed ion beam (Cs or microfocused Ga) to remove molecules from the very outermost surface of the sample. The particles are removed from atomic monolayers on the surface (secondary ions). These particles are then accelerated into a "flight tube" and their mass is determined by measuring the exact time at which they reach the detector (i.e. time-of-flight). Three operational modes are available using ToF-SIMS: surface spectroscopy, surface imaging and depth profiling. Analytical capabilities of ToF-SIMS include:

- Mass resolution of 0.00x amu. Particles with the same nominal mass (e.g. Si and C<sub>2</sub>H<sub>4</sub>, both with amu = 28) are easily distinguished from one another because as Mr. Einstein predicted there is a slight mass shift as atoms enter a bound state.
- Mass range of 0-10,000 amu; ions (positive or negative), isotopes, and molecular compounds (including polymers, organic compounds, and up to ~amino acids) can be detected.
- Trace element detection limits in the ppm range.
- Sub-micron imaging to map any mass number of interest.
- Depth profiling capabilities; sequential sputtering of surfaces allow analysis of the chemical stratigraphy on material surfaces (typical sputtering rates are ~100 Å/minute).
- Retrospective analysis. Every pixel of a ToF-SIMS map represents a full mass spectrum. This allows an analyst to retrospectively produce maps for any mass of interest, and to interrogate regions of interest (ROI) for their chemical composition via computer processing after the dataset has been instrumentally acquired.

### Fundamental Principles of Time-of-Flight Secondary Ion Mass Spectrometry (ToF-SIMS)

ToF-SIMS uses a focused, pulsed particle beam (typically Cs or Ga) to dislodge chemical species on a materials surface. Particles produced closer to the site of impact tend to be dissociated ions (positive or negative). Secondary particles generated farther from the impact site tend to be molecular compounds, typically fragments of much larger organic macromolecules. The particles are then accelerated into a flight path on their way towards a detector.

Because it is possible to measure the "time-of-flight" of the particles from the time of impact to detector on a scale of nano-seconds, it is possible to produce a mass resolution as fine as 0.00X atomic mass units (i.e. one part in a thousand of the mass of a proton). Under typical operating conditions, the results of ToF-SIMS analysis include:

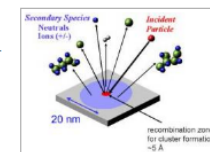
- a mass spectrum that surveys all atomic masses over a range of 0-10,000 amu,
- depth profiles are produced by removal of surface layers by sputtering under the ion beam.

ToF-SIMS is also referred to as "static" SIMS because a low primary ion current is used to "tickle" the sample surface to liberate ions, molecules and molecular clusters for analysis. In contrast, "dynamic" SIMS is the method of choice for quantitative analysis because a higher primary ion current results in a faster sputtering rate and produces a much higher ion yield. Thus, dynamic SIMS creates better counting statistics for trace elements. Organic compounds are effectively destroyed by "dynamic" SIMS, and no diagnostic information is obtained.

### Time-of-Flight Secondary Ion Mass Spectrometry (ToF-SIMS) Instrumentation – How Does It Work?

ToF-SIMS instruments typically include the following components:

- An ultrahigh vacuum system, which is needed to increase the mean free path of ions liberated in the



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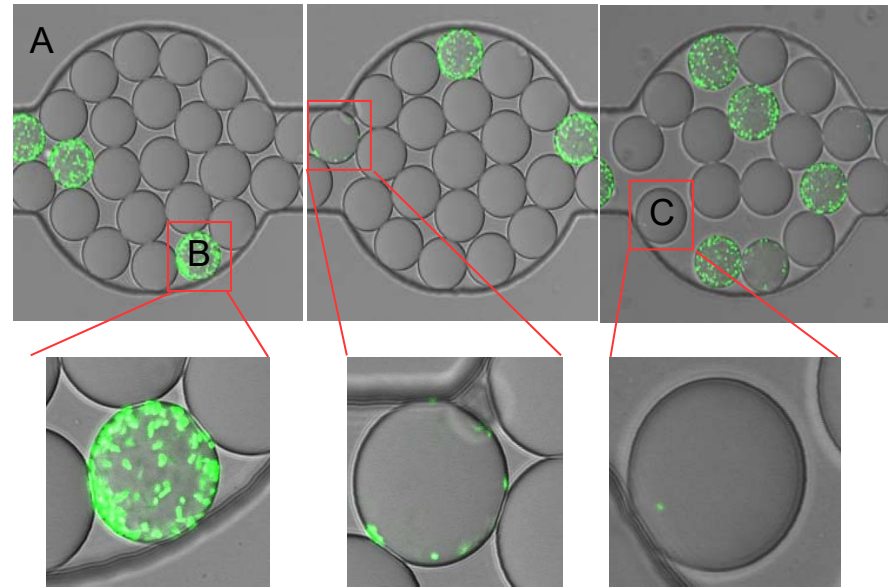
# Research Highlights - Microfluidics

## “Droplet Engineering” to Study Single Bacteria Cells



Prof. Connie Chang  
Chemical and  
Biological Engineering,  
Center for Biofilm  
Engineering

C.B. Chang, J.N. Wilking, S.H. Kim, H.C. Shum, D. A. Weitz. “Monodisperse Emulsion Drop Microenvironments for Bacterial Biofilm Growth” *Small* (2015)



Examples of cell resuscitation of individual *P. aeruginosa* bacteria within 15 μm diameter oil microdroplets.  
Maximum Projection images from CLSM.



# Research Highlights – Nonlinear Optics

Industrial user AdvR., Inc. of Bozeman uses MONT to build laser wavelength converters

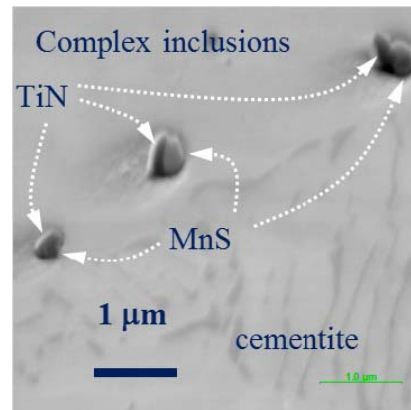
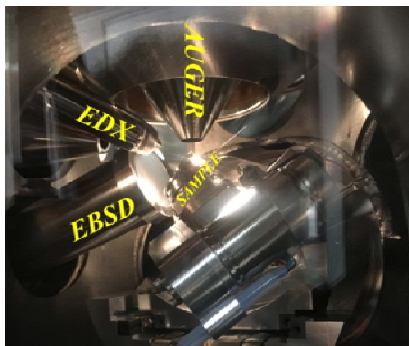


AdvR uses the MMF/MONT facilities to fabricate waveguides in nonlinear optical materials such, as KTP, LN, and Mg:LN.

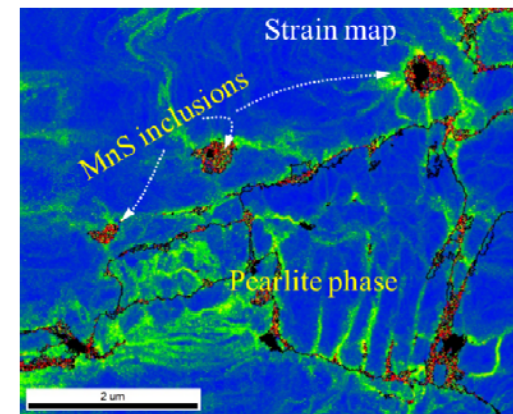
These waveguides can be periodically poled in various configurations and used to generate second harmonic generation, sum frequency generation, downconversion, difference frequency generation, Bragg stabilization, and phase modulation.

# Research Highlights – PHI 710 NanoAuger

Hybrid Auger system combines a field emission Auger nanoprobe with an EDX and an EBSD system. This enables analysis of morphology, surface and bulk composition, structural, crystallographic orientation and phase at the same area at the nanoscale resolution.



SEM image of MnS, TiN inclusions and cementite lamella in carbon steel.



Overlaid strain map of the same region obtained using EBSD.

The role of nano-defects in the localized corrosion of low carbon steel (a USD 1.1 Trillion annual problem).

# MONT Had a Successful NNCI Y1!

- Significant facility enhancement and new hires; excellent leveraging of NSF funds to grow State/Private investment
- Growth in user numbers and diversity
- Strong E & O portfolio, locally and on the web in partnership with Carleton College's SERC
- Exciting new research directions, both academic and industrial